

中药复方及其有效成分对降低 免疫检查点抑制剂耐药性的研究进展

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摘要:严重的耐药性是免疫检查点抑制剂(immune checkpoint inhibitors, ICIs)急需攻克的难题。目前为止,现代医学对此还没有很好的解决办法。中医药普遍具有毒性低、不良反应小、疗效显著的特点,与现代医学联合,在肿瘤治疗的各个方面都发挥着积极作用。因此,可以尝试从中医药领域寻找突破,提高 ICIs 的疗效,达到增效减毒的目的。回顾了近年来对 ICIs 耐药性发生机制的最新研究,以及中医药对逆转其耐药性的突破性进展,并总结讨论,以期能够为中医药缓解 ICIs 耐药性方面研究提供帮助。

关键词:中医药;免疫检查点抑制剂;耐药性;免疫

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Research Progress on Reducing Resistance of Immune Checkpoint Inhibitors with Traditional Chinese Medicine

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Abstract: The resistance to immune checkpoint inhibitors (ICIs) presents a critical challenge that necessitates immediate attention. Currently, modern medicine lacks effective solutions for this issue. Traditional Chinese medicine (TCM), on the other hand, is known for its attributes of low toxicity, minimal side effects and notable therapeutic efficacy. When integrated with modern medicine, TCM demonstrates a positive role across various aspects of tumor treatment. Consequently, exploring potential breakthroughs in the realm of TCM becomes a worthwhile pursuit to enhance the effectiveness of ICIs while minimizing their toxicity. This review article aimed to outline the recent advancements in understanding the mechanisms underlying ICIs resistance and highlighted the groundbreaking progress achieved by TCM in overcoming this resistance. Through comprehensive analysis and synthesis of the existing research, this review aimed to contribute valuable insights to TCM potential in mitigating resistance to ICIs.

Keywords: traditional Chinese medicine; immune checkpoint inhibitors; drug resistance; immunity

免疫治疗是近年来兴起的新型抗肿瘤方式,尤以免疫检查点抑制剂(immune checkpoint inhibitors, ICIs)起到了里程碑式作用^[1-5]。ICIs 主要包括细胞毒性 T 淋巴细胞相关抗原-4 (cytotoxic T lymphocyte-associated antigen-4, CTLA-4) 抑制

剂、程序性细胞死亡蛋白-1 (programmed cell death-1, PD-1) 抑制剂及细胞程序性死亡配体-1 (programmed cell death ligand-1, PD-L1) 抑制剂。近年来,又发现了淋巴细胞活化基因-3 (Lymphocyte activation gene 3, LAG-3)、T 细胞免疫球蛋白黏蛋白分子-3 (T-cell immunoglobulin mucin molecule-3, TIM-3)、T 细胞免疫球蛋白与免疫受体酪氨酸抑制基序结构域 (T Cell Immunoglobulin and Immunoreceptor Tyrosine-based Inhibition Motif Domain, TIGIT)、CD276、T 细胞激活抑制物免疫球蛋白可变区结构域 (V-domain Ig Suppressor of T Cell Activation, VISTA) 等新的免疫检查点,有望研发出新的抑制剂应用于临床^[6-7]。

ICIs 多为单克隆抗体,具有显著的临床疗效和特异性,但是,也会在肝细胞癌、微卫星稳定型结直肠癌、激素受体阳性乳

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腺癌等诸多癌症中,引发严重的免疫耐药^[8-10],大大影响药物疗效,使许多患者无法从中获益。ICIs 有关的耐药性根据发生时间可分为:(1)原发性耐药:指 ICIs 从开始就无法激活人体对肿瘤的免疫应答;(2)适应性耐药:指免疫系统可以识别肿瘤,但肿瘤为躲避识别,产生免疫逃逸,导致耐药性;(3)获得性耐药:指 ICIs 治疗一段时间后,出现耐药性。也可以分为肿瘤自身引起的内源性耐药性和肿瘤微环境引起的外源性耐药性。

对于 ICIs 治疗中出现的免疫耐药,现代医学通常选择联合用药的方式来缓解患者耐药的情况。近年,癌症免疫治疗协会针对不同分期的肿瘤患者,利用 ICIs 联合化疗、靶向治疗来解决免疫耐药的治疗手段,提供了最新的指导方针^[11-13]。同时,部分研究表明,放疗可通过重塑肿瘤微环境(tumor micro-environment, TME)缓解肿瘤的耐药性状态^[14-15]。但联合治疗的使用,会极大增加患者出现诸多严重免疫相关不良反应(immune-related adverse events, IRAES)的风险^[16-19]。因此,凭借中医药性平和,不良反应小的优势,可以尝试从中医药领域干预 ICIs 产生的耐药性。

本文对 ICIs 的作用机制、耐药性机制以及中医药对 ICIs 耐药性的干预作用进行总结讨论,并提出自己的观点,为科研人员提供新的研究思路,希望可以借助中医药的手段,解决 ICIs 耐药性方面的难题,达到中西医结合免疫治疗肿瘤的最佳效果。

1 ICIs 的作用机制

ICIs 的作用机制现已趋于完善,总体而言,就是通过解除负性共刺激分子 CTLA-4 和 PD-1/PD-L1 对于 T 细胞的抑制作用,防止活化的 T 细胞失活和肿瘤细胞逃逸,使 T 细胞能够无限制地杀伤肿瘤细胞^[20-28]。如插页 IX 图 1。

2 引起 ICIs 耐药性的机制

目前,已知引起 ICIs 耐药性的机制主要有:

(1) 肿瘤表面新抗原的缺失:①肿瘤突变负荷的降低^[29-32];②对含有新抗原转录本的抑制^[33];③基因拷贝数丢失^[5,33];④克隆选择和表观遗传抑制^[5,29-30,34];⑤微卫星不稳定性的减弱^[29,35-37]。

(2) 抗原提呈机制的缺陷:①β2-微球蛋白基因和人类白细胞抗原的丢失^[5,29-30];②γ-干扰素(interferon-gamma, IFN-γ)、α-干扰素(interferon alfa, IFN-α)信号通路的异常^[5,30-32];③树突状细胞(dendritic cells, DCs)的功能异常^[30,38];④主要组织相容性复合体-I类分子(major histocompatibility complex-I, MHC-I)的基因、转录异常^[31]。

(3) 肿瘤介导的免疫抑制:①磷酸酯酶与张力蛋白同源物(phosphatase and tensin homolog, PTEN)的缺失^[39];②WNT/β-catenin 的活性减弱^[40]。

(4) TME 的改变:①抑制上调 T 细胞转运的趋化因子的表达^[16,30];②阻止 T 细胞浸润^[16,30];③免疫抑制细胞的大量增殖^[16,30];④对 T 细胞的物质供应和代谢改变^[16,41];⑤肿瘤细胞外囊泡^[42]。

(5) PD-L1 的表达上调:①相关基因突变^[30];②抑癌基

因失活^[30];③免疫抑制因子的释放^[30]。

(6) 其他抑制性免疫检查点的上调(如 LAG-3、TIGIT、B 及 T 淋巴细胞衰减因子、TIM-3)^[9,16,30]。

(7) 体内肠道微生物的影响^[16,37]。

而更多的耐药性机制还在陆续被发现。近年,吕志民团队^[43]又发现了新的耐药机制,突变的表皮因子生长受体可提高肿瘤细胞中 CD55 和 CD59 的表达,从而抑制补体的激活和 CD₈⁺T 细胞的活性。此外,导致 ICIs 耐药的部分原因是微环境中缺乏肿瘤浸润淋巴细胞(tumor-infiltrating lymphocytes, TIL),最新研究发现,铁死亡、细胞焦亡、自噬和坏死性凋亡等调节性细胞死亡,可改变 TME 和 TIL 的流入^[44],与调节 ICIs 耐药性方面有着密切联系。

KEISUKE YAMAMOTO^[45]指出,自噬抑制剂可通过恢复 T 细胞表面 MHC-I 的表达,重建抗原呈递机制,增强 T 细胞抗肿瘤免疫反应和抑制肿瘤细胞的免疫逃逸,并且发现,氯喹自噬抑制剂与双重 ICIs(抗 PD-1 和抗 CTLA-4 抗体)一起使用,可大大增强胰腺导管腺癌对 ICIs 的敏感性,但值得注意的是,氯喹自噬抑制剂与 PD-1 抗体单独使用,效果甚微。人黑色素瘤中,自噬标志物——微管关联蛋白及其他自噬体可降低黑色素瘤抗原-A(melanoma antigen-associated gene-A, MAGE-A)和三重基序蛋白 28(tripartite motif-containing 28, TRIM28)的表达,二者都可引起肿瘤对 CTLA-4 的耐药,这表明不仅是自噬抑制剂,自噬诱导剂(如脂联素,2-氨基烟腈类化合物)与 CTLA-4 抑制剂也可协同治疗肿瘤^[46-50]。

铁死亡可以限制免疫抑制细胞的功能,并有助于改善机体对免疫治疗的原发性耐药,提高 ICIs 的疗效^[51-57]。也可以与其他细胞因子相互调节,间接提高肿瘤对 ICIs 的敏感性,或者代偿性地抑制 CTLA-4、PD-1/PD-L1 活性。之前已有研究证实,IFN-γ 可通过促进 MHC-I 的表达和 CD₈⁺T 细胞的激活来降低机体对 ICIs 的耐药性^[58-61],最近又有大量研究肯定了 CD₈⁺T 细胞可通过分泌 IFN-γ 介导肿瘤细胞对铁死亡的敏感性^[62-65]。2019 年,WANG 及其同事^[62]首次发现了 IFN-γ 在降低溶质载体家族 3 成员 2 和溶质载体家族 7 成员 11 蛋白表达从而增强铁死亡的重要性。2022 年,又有研究团队发现了花生四烯酸(AA)结合 IFN-γ 可诱导肿瘤铁死亡^[63]。综上所述,IFN-γ、铁死亡、ICIs 三者之间联系密切,IFN-γ 可能通过铁死亡途径降低机体对 ICIs 的耐药性。

造成 ICIs 耐受的一个重要原因是肿瘤微环境中缺乏 TIL,此类肿瘤称为“冷”肿瘤。WANG 等^[66]将接种 4T1“冷”肿瘤的小鼠分为两组,一组使用 NP-GSDMA3 和三氟硼酸基苯丙氨酸,另一组使用 PD-1。治疗一轮后,两组均无明显疗效,但三种药物联合使用时,肿瘤明显受到抑制。除此之外,KOJI HARATANI^[67]、DAN A ERKES^[68]也都做了相关研究,得出类似结论。这证明 ICIs 只有在发生细胞焦亡的情况下才能杀死“冷”肿瘤细胞,并且单一的细胞焦亡并不能有效地抑制肿瘤,更加强调了焦亡诱导剂与 ICIs 联合使用治疗“冷”肿瘤的重要性,但二者之间产生影响的具体机制,相关研究甚少,可能与诱导炎症细胞死亡的机制有关^[69]。

最新发现了 WNT/ β -连环蛋白信号通路会通过抑制趋化因子 C-C 配体 4、DCs 的聚集等影响 T 细胞的招募,诱导非 T 细胞浸润到 TME,也可以通过与肿瘤相关巨噬细胞的相互作用来调节 TME,或促进乳酸的产生,抑制细胞毒性 T 淋巴细胞的产生,进而削弱 PD-1/PD-L1 抑制剂的治疗效果^[70]。

3 中药复方及其有效成分对 ICI 耐药性的干预作用

3.1 青蒿琥酯 (artesunate, ART) ART 是从中药青蒿中提取的一种重要的青蒿素衍生物,具有显著的抗肿瘤活性和低毒性^[71-72]。PDZ 结合基序的转录共激活因子 (PDZ-binding motif, TAZ), 可通过调节 PD-L1 介导肿瘤细胞的免疫耐药^[73], 国内一项研究首次发现了 ART 靶向 TAZ-TEAD 复合物, 其疏水基团与 TAZ-TEAD 复合物的疏水口袋紧密结合以蛋白酶体依赖性方式促进 TAZ 降解, 还可以通过募集浸润性 CD₈⁺ T 细胞, 克服非小细胞肺癌对 ICI 的耐药性^[72]。同时, ART 具有不错的抗肿瘤作用, 能够通过下调谷胱甘肽过氧化物酶 4 来诱导铁死亡, 通过降低 TAZ/PD-L1 对 T 细胞的生长抑制, 促进癌细胞死亡^[72,74]。可以对 ART 作用机制、安全性等方面做更深入研究, 望作为一款全新的抗肿瘤药物早日应用临床。

3.2 β -榄香烯 β -榄香烯是从温郁金中提取的活性成分提取物, 可以抑制肿瘤增殖, 调节免疫系统, 降低 P-糖蛋白的表达, 从而逆转多药耐药 (multidrug resistance, MDR)^[75-76]。近年, CHEN P^[77] 首次发现 β -榄香烯是一种新的铁死亡诱导剂, 与西妥昔单抗联合使用可促进诱导铁死亡, 从而加强对 KRAS 突变的结直肠癌细胞的敏感性, 也可通过 Pole2 介导的 p53 和 PI3K/AKT 信号诱导肺癌细胞发生铁死亡^[78]。PTEN 的缺失是 ICI 耐药的一个重要原因, 最近研究发现, β -榄香烯可以通过 METTL3 介导的 N⁶ 甲基腺苷修饰促进 PTEN 的表达, 但未对具体的作用机制作深入研究^[79]。除此之外, β -榄香烯还可以通过调节 AMPK/MAPK、JAK2/STAT3、PI3K/AKT 等信号通路逆转 MDR^[79-80]。

3.3 高良姜素 (galangin, GAL) 已知 GAL 是从高良姜中提取的具有抗增殖、抗炎、促凋亡、抑制血管生成和转移的中药提取物, 具有良好的抗肿瘤作用。之前已有学者发现 GAL 也能够促进 PTEN 的表达^[81-82], 最近, 韩立卓^[83] 做了相关机制研究得出结论, GAL 通过抑制 JAK1/JAK2/Src 和 Ras/RAF/MEK/ERK 信号通路抑制 STAT3 和 Myc 的激活, 并通过抑制 STAT3 和 Myc 之间的相互作用来抑制肿瘤表面 PD-L1 蛋白表达, 降低肿瘤表面 PD-L1 与 T 细胞表面 PD-1 结合, 恢复 T 淋巴细胞的活力和对肿瘤细胞的特异性杀伤力。有报道还发现了 GAL 可调节 PI3K/AKT/mTOR 信号通路抑制血管损伤诱导的新内膜形成^[84], 通过抑制 MAPK 和 NF- κ B 信号通路抑制 NF- κ B 配体激活因子诱导的破骨形成^[85]。PD-L1 的表达、T 淋巴细胞活力和 STAT 蛋白家族都是导致 ICI 耐药的重要因素, 且与 JAK、PI3K/AKT、MAPK 和 NF- κ B 等信号通路有密切关系。

3.4 人参多糖 扶正类中药可以通过调节肿瘤微环境的免疫抑制状态, 增加 CD₈⁺ T 细胞的浸润, 控制肿瘤细胞的生长^[86],

人参中的有效成分人参多糖, 可以提高机体 TNF- α 的水平, 研究发现, 以人参为君药的四君子汤单独或与 STAT3 抑制剂联合使用可降低 PD-1、PD-L1、STAT3 的表达, 增强 NK 细胞对小鼠结肠癌皮下瘤的杀伤作用^[87]。肿瘤细胞常通过高表达 FasL, 抑制 Fas 来诱导效应 T 细胞凋亡, 促进免疫逃逸, 含人参成分的养胃抗癌方则可以逆转这一情况, 并且还能增强肿瘤细胞对 Fas/Fas L 凋亡通路的敏感性^[88]。近年, 有学者将接种 Lewis 肺癌和 B16-F10 细胞的 C57BL/6J 小鼠和人源化 PD-1 敲入小鼠注射 α PD-1 单克隆抗体和人参多糖, 结果发现人参多糖可以通过改变肠道微生物群和色氨酸/色氨酸的比例, 增强抗 PD-1/PD-L1 免疫治疗的抗肿瘤作用^[89]。

3.5 黄芪多糖 (astragalus polysaccharin, APS) APS 灌胃的 Lewis 肺癌小鼠 CD₄⁺ T 细胞及 CD4/CD8T 细胞的比率显著提升, ELISA 测定小鼠血清中 IFN- γ 和 IL-2 含量都明显升高, 免疫印迹结果显示 APS 还可以降低肿瘤组织中 PD-L1 的表达量^[90]。APS 还可以作用于 IL-6/STAT3、TNF- α /NF- κ B 等通路, 促进骨髓间充质干细胞向肿瘤相关成纤维细胞分化, 调节脂质代谢等^[91-92]。CHANG 等人^[93] 通过体内、体外实验, 也证实了 APS 在肿瘤细胞可降低 PD-L1 的表面表达和蛋白质水平, 能够有效地克服肿瘤免疫逃避及其免疫耐受。

3.6 白术多糖 白术多糖已被证明具有良好的促免疫作用, 人们通过 miRNA 测序发现, 白术多糖可能是通过 novel-mir2 靶向 CTLA-4 上调 TCR-NFAT 途径来减轻免疫抑制^[94]。此外, PD-L1 为 miR-34a 的靶基因, 韩懿存^[95] 通过转染 miR-34a mimics 48 h 后发现, 过表达的 miR-34a 可以抑制 PD-L1 mRNA 的表达水平, 白术多糖能够通过促进 miR-34a 的过表达, 来降低 PD-L1 的表达, 不仅可以缓解 ICI 的耐药性, 还有望成为一款重要的抗肿瘤单体。

3.7 鸦胆子苦醇 (brusatol, BRU) NRF2 信号通路表达的降低, 可显著诱导 CD₈⁺ 和 CD₄⁺ T 细胞浸润肿瘤, 以抑制黑色素瘤进展, 还能够减弱 PD-L1 的表达^[17]。BRU 能够增强 NRF2 的泛素化和促进其蛋白的降解, 有效抑制 NRF2 通路的活性^[18-19,96], 现已成为一种公认的 NRF2 抑制剂。同时, BRU 还可以通过调控 PTEN/PI3K/AKT 等信号通路^[97], 抑制原癌基因从而促进缺氧诱导因子-1 α 的降解等方式^[98], 达到抑制肿瘤生长的目的。部分学者指出, BRU 与 ICI 的联合使用, 在改善机体免疫耐受的同时, 还可以增强抗肿瘤的功效^[17,99]。

3.8 姜黄素 姜黄素是中药姜黄的主要活性成分, 具有重要的免疫调节作用。有学者曾对小鼠荷瘤模型联合使用姜黄素和免疫检查点阻断治疗, 结果发现, 姜黄素通过抑制 DCs 和癌细胞中的 STAT3 信号通路, 来增强肿瘤抗原特异性 T 细胞的表达, 并且姜黄素和 PD-1/PD-L1 抑制剂的联合使用, 具有显著的抗肿瘤作用, IL-6 高表达的癌症患者^[100] 尤为突出。有研究发现, 姜黄素与 PD-1 抑制剂的协同作用, 可减缓 Hep3B 细胞增殖, 激活淋巴细胞, 抑制免疫逃避, 并下调转化生长因子- β 1 表达^[101]。

3.9 楝酰胺 (roglamide, RocA) RocA 是从米仔兰植物中提取的一味中药单体, 可以通过抑制自噬和激活 cGAS-STING

信号通路促进NK细胞的浸润和抗肿瘤免疫^[102-103],RocA能够将“冷”肿瘤变为“热”肿瘤,促进肿瘤浸润T细胞的增殖,与PD-1抑制剂可共同降低CD4⁺T细胞中PD-1表达水平^[104],大大增强了免疫治疗的疗效。

3.10 槲皮素 槲皮素是一种广泛存在于侧柏叶、姜黄、补骨脂等多种中药中的黄酮醇类化合物,可逆转许多细胞的多重耐药性^[105],通过HCV,HSV-1,H1N1,HBV,SARS-CoV2和HIV-1等信号通路调节人体免疫功能。研究表明,槲皮素可以通过抑制PD-1/PD-L1间的相互作用,减弱PD-L1对T细胞的抑制作用,还能够提高CD8,颗粒酶B和IFN- γ 等细胞分子的表达^[106]。

3.11 葛根芩连汤(Gegen Qinlian decoction,GQD) 目前,GQD对于结直肠癌有良好的治疗效果。对于微卫星稳定型结直肠癌,免疫检查点抑制剂的治疗效果甚微,存在严重的免疫耐药现象。GQD则能够通过控制Wnt/ β -catenin信号通路,调节鞘磷脂和甘油磷脂的脂质代谢途径,提高免疫检查点抑制剂的抗肿瘤活性,遏制肿瘤细胞的免疫逃逸,和PD-1抑制剂协同使用可调节肠道微生物菌群,重塑肿瘤微环境,增加CT8⁺T细胞、IFN- γ 的表达,此外,二者在下调PD-1表达的同时,还提高了IL-2的水平^[107-109]。二者的联合使用可以成为微卫星稳定型结直肠癌患者的一种新型治疗策略。

3.12 健脾滋肾泻火方/健脾滋肾颗粒 健脾滋肾泻火方现常用于改善原发性免疫性血小板减少症。上海中医药大学李晓靖^[110]近期探究了其中的作用机制,结果发现,10%健脾滋肾泻火方20g/kg含药血清组中JAK1、STAT1的mRNA和蛋白表达均明显下调,健脾滋肾泻火方也是通过抑制JAK/STAT信号通路对ITP起到治疗效果;临床研究中发现,中成药健脾滋肾颗粒可以影响补体调节蛋白CD55和CD59的表达^[111],这些都是改善ICIs耐药的重要通路。

4 总结与展望

ICIs在肿瘤治疗方面疗效明显,但因其引起的耐药性使许多患者无法从中受益。中医药在此方面已取得许多突破性进展,拥有广阔的研究前景。作者认为:(1)可以通过网络药理学,生物信息学分析等数据挖掘的方式,挖掘出其他未知的、导致免疫逃逸的信号通路,以及能够调控相关通路的单药或者复方;(2)目前大部分开展的是中药与ICIs的基础研究,缺乏临床证据,对于像GQD这类经方,可以尽早应用于临床研究;(3)基于中医基础理论,从理论角度出发,深入探索中医药与免疫治疗之间的联系,拓宽思路,找到更多能够抑制免疫逃逸的药物;(4)目前的现代医学临床疗效评价标准是否适合中医药联合ICIs的治疗有待商榷,需要制订符合中医药联合ICIs的现代医学临床疗效评价标准,能够使用药规范、有据可依。

参考文献

[1] YU J H, YIN Y C, YU Y, et al. Effect of concomitant antibiotics use on patient outcomes and adverse effects in patients treated with ICIs [J]. Immunopharmacology And Immunotoxicology, 2023, 45 (3): 386-394.
[2] U DAFNI, Z TSOURTI, K VERVITA, et al. Immune checkpoint in-

hibitors, alone or in combination with chemotherapy, as first-line treatment for advanced non-small cell lung cancer. A systematic review and network meta-analysis [J]. Lung Cancer, 2019, 134: 127-140.
[3] VAF AEI S, ZEKIY A O, KHANAMIR R A, et al. Combination therapy with immune checkpoint inhibitors (ICIs), a new frontier [J]. Cancer Cell International, 2022, 22(1):2.
[4] SONG Y X, FU Y, XIE Q, et al. Anti-angiogenic agents in combination with immune checkpoint inhibitors: a promising strategy for cancer treatment [J]. Front Immunol, 2020, 11: 1956.
[5] SCHOENFELD A J, HELLMANN M D. Acquired resistance to immune checkpoint inhibitors [J]. Cancer Cell, 2020, 37(4): 443-455.
[6] QIN S, XU L P, YI M, et al. Novel immune checkpoint targets: moving beyond PD-1 and CTLA-4 [J]. Mol Cancer, 2019, 18(1): 155.
[7] HUANG X, ZHANG X Z, LI E I, et al. VISTA: an immune regulatory protein checking tumor and immune cells in cancer immunotherapy [J]. J Hematol Oncol, 2020, 13(1): 83.
[8] WANG Z J, WANG Y C, GAO P. Immune checkpoint inhibitor resistance in hepatocellular carcinoma [J]. Cancer Lett, 2023, 555:216038.
[9] VATHIOTIS I A, TRONZAS I, GAVRIELATOU N, et al. Immune checkpoint blockade in hormone receptor-positive breast cancer: resistance mechanisms and future perspectives [J]. Clin Breast Cancer, 2022, 22(7):642-649.
[10] FANG Y H, SUN H Y, XIAO X H, et al. Low-dose immunogenic chemotherapeutics promotes immune checkpoint blockade in microsatellite stability colon cancer [J]. Front Immunol, 2022, 13:1040256.
[11] RIZVI N, ADEMUYIWA F O, CAO Z A, et al. Society for Immunotherapy of Cancer (SITC) consensus definitions for resistance to combinations of immune checkpoint inhibitors with chemotherapy [J]. J Immunother Cancer, 2023, 11(3):e005920.
[12] ATKINS M B, ASCIERTO P A, FELTQUATE D, et al. Society for immunotherapy of cancer (SITC) consensus definitions for resistance to combinations of immune checkpoint inhibitors with targeted therapies [J]. J Immunother Cancer, 2023, 11(3):e005923.
[13] KLUGER H, BARRETT J C, GAINOR J F, et al. Society for Immunotherapy of Cancer (SITC) consensus definitions for resistance to combinations of immune checkpoint inhibitors [J]. J Immunother Cancer, 2023, 11(3):e005921.
[14] PENG J F, YIN X Y, YUN W H, et al. Radiotherapy-induced tumor physical microenvironment remodeling to overcome immunotherapy resistance [J]. Cancer Lett, 2023, 559:216108.
[15] LAN YAN, MOUSTAFA MAHMOUD, KNOLL MAXIMILIAN, et al. Simultaneous targeting of TGF- β /PD-L1 synergizes with radiotherapy by reprogramming the tumor microenvironment to overcome immune evasion [J]. Cancer Cell, 2021, 39(10):1388-1403. e10.
[16] WANG Z Y, WU X Y. Study and analysis of antitumor resistance mechanism of PD1/PD-L1 immune checkpoint blocker [J]. Canc-

- er Med, 2020, 9(21): 8086 – 8121.
- [17] ZHU B, TANG L M, CHEN S Y, et al. Targeting the upstream transcriptional activator of PD – L1 as an alternative strategy in melanoma therapy[J]. *Oncogene*, 2018, 37(36): 4941 – 4954.
- [18] CAI S J, LIU Y, HAN – SUE. Brusatol, an NRF2 inhibitor for future cancer therapeutic[J]. *Cell Biosci*, 2019, 9: 45.
- [19] CHENG C, YUAN F, CHEN X P, et al. Inhibition of Nrf2 – mediated glucose metabolism by brusatol synergistically sensitizes acute myeloid leukemia to Ara – C [J]. *Biomed Pharmacother*, 2021, 142: 111652.
- [20] WEI S C, DUFFY C R, ALLISON J P. Fundamental mechanisms of immune checkpoint blockade therapy[J]. *Cancer Discov*, 2018, 8(9): 1069 – 1086.
- [21] SUGIURA D, MARUHASHI T, OKAZAKI I M, et al. Restriction of PD – 1 function by cis – PD – L1/CD80 interactions is required for optimal T cell responses [J]. *Science*, 2019, 364 (6440): 558 – 566.
- [22] KUMAR A, CHAMOTO K, CHOWDHURY P S, et al. Tumors attenuating the mitochondrial activity in T cells escape from PD – 1 blockade therapy[J]. *Elife*, 2020, 9: e52330.
- [23] ROWSHANRAVAN B, HALLIDAY N, SANSOM D M. CTLA – 4: a moving target in immunotherapy[J]. *Blood*, 2018, 131(1): 58 – 67.
- [24] ROTTE A. Combination of CTLA – 4 and PD – 1 blockers for treatment of cancer[J]. *J Exp Clin Cancer Res*, 2019, 38(1): 255.
- [25] PAUKEN K E, TORCHIA J A, CHAUDHRI A, et al. Emerging concepts in PD – 1 checkpoint biology[J]. *Semin Immunol*, 2021, 52: 101480.
- [26] NISHIMURA C D, PULANCO M C, CUI W, et al. PD – L1 and B7 – 1 cis – interaction: new mechanisms in immune checkpoints and immunotherapies [J]. *Trends Mol Med*, 2021, 27 (3): 207 – 219.
- [27] HURRELL B P, HELOU D G, HOWARD E, et al. PD – L2 controls peripherally induced regulatory T cells by maintaining metabolic activity and Foxp3 stability[J]. *Nat Commun*, 2022, 13(1): 5118.
- [28] HUI E F, CHEUNG JEANNE, ZHU J, et al. T cell costimulatory receptor CD28 is a primary target for PD – 1 – mediated inhibition [J]. *Science*, 2017, 355(6332): 1428 – 1433.
- [29] BAGCHI S, YUAN R, ENGLEMAN E G. Immune checkpoint inhibitors for the treatment of cancer: clinical impact and mechanisms of response and resistance [J]. *Annu Rev Pathol*, 2021, 16: 223 – 249.
- [30] SUN Q Y, WEI X Z, WANG Z L, et al. Primary and acquired resistance against immune check inhibitors in non – small cell lung cancer[J]. *Cancers (Basel)*, 2022, 14(14): 3294.
- [31] VEGA D M, YEE L M, MCSHANE L M, et al. Aligning tumor mutational burden (TMB) quantification across diagnostic platforms: phase II of the Friends of Cancer Research TMB Harmonization Project[J]. *Ann Oncol*, 2021, 32(12): 1626 – 1636.
- [32] DE LORENZO S, TOVOLI F, TREVISANI F. Mechanisms of primary and acquired resistance to immune checkpoint inhibitors in patients with hepatocellular carcinoma[J]. *Cancers (Basel)*, 2022, 14(19): 4616.
- [33] ROSENTHAL R, CADIEUX E L, SALGADO R, et al. Neoantigen – directed immune escape in lung cancer evolution [J]. *Nature*, 2019, 567(7749): 479 – 485.
- [34] CHEN S Q, XIE P, COWAN M, et al. Epigenetic priming enhances antitumor immunity in platinum – resistant ovarian cancer [J]. *J Clin Invest*, 2022, 132(14): e158800.
- [35] LU C Z, GUAN J H, LU STEVE, et al. DNA sensing in mismatch repair – deficient tumor cells is essential for anti – tumor immunity [J]. *Cancer Cell*, 2021, 39(1): 96 – 108. e6.
- [36] ROUDKO V, BOZKUS C C, ORFANELLI T, et al. Shared immunogenic poly – epitope frameshift mutations in microsatellite unstable tumors[J]. *Cell*, 2020, 183(6): 1634 – 1649. e17.
- [37] ROBERTO M, CARCONI C, CERRETI M, et al. The challenge of ICIs resistance in solid tumours: could microbiota and its diversity be our secret weapon? [J]. *Front Immunol*, 2021, 12: 704942.
- [38] MARTIN K, SCHREINER J, ZIPPELIUS A. Modulation of apc function and anti – tumor immunity by anti – cancer drugs [J]. *Front Immunol*, 2015, 6: 501.
- [39] ÁLVAREZ – GARCIA V, TAWIL Y, WISE H M, et al. Mechanisms of PTEN loss in cancer: It’s all about diversity[J]. *Semin Cancer Biol*, 2019, 59: 66 – 79.
- [40] LIU J Q, XIAO Q, XIAO J N, et al. Wnt/ β – catenin signalling: function, biological mechanisms, and therapeutic opportunities [J]. *Signal Transduct Target Ther*, 2022, 7(1): 3.
- [41] MUNN D H, MELLOR A L. IDO in the tumor microenvironment: inflammation, counter – regulation, and tolerance [J]. *Trends Immunol*, 2016, 37(3): 193 – 207.
- [42] CHEN J M, YANG J, WANG W H, et al. Tumor extracellular vesicles mediate anti – PD – L1 therapy resistance by decoying anti – PD – L1 [J]. *Cell Mol Immunol*, 2022, 19(11): 1290 – 1301.
- [43] SHAO F, GAO Y B, WANG W, et al. Silencing EGFR – upregulated expression of CD55 and CD59 activates the complement system and sensitizes lung cancer to checkpoint blockade [J]. *Nat Cancer*, 2022, 3(10): 1192 – 1210.
- [44] NIU X, CHEN L J, LI Y, et al. Ferroptosis, necroptosis, and pyroptosis in the tumor microenvironment: Perspectives for immunotherapy of SCLC [J]. *Semin Cancer Biol*, 2022, 86(Pt 3): 273 – 285.
- [45] YAMAMOTO K, VENIDA A, YANO J, et al. Autophagy promotes immune evasion of pancreatic cancer by degrading MHC – 1 [J]. *Nature*, 2020, 581(7806): 100 – 105.
- [46] SHUKLA S A, BACHIREDDY P, SCHILLING B, et al. Cancer – germline antigen expression discriminates clinical outcome to CTLA – 4 blockade [J]. *Cell*, 2018, 173(3): 624 – 633. e8.
- [47] LIN J H, GUO D J, LIU H, et al. The SETDB1 – TRIM28 complex suppresses antitumor immunity [J]. *Cancer Immunol Res*, 2021, 9(12): 1413 – 1424.
- [48] CHUNG S J, NAGARAJU G P, NAGALINGAM A, et al. ADIPOQ/adiponectin induces cytotoxic autophagy in breast cancer cells through STK11/LKB1 – mediated activation of the AMPK – ULK1 axis [J]. *Autophagy*, 2017, 13(8): 1386 – 1403.
- [49] ZHANG P H, ZHENG Z G, LING L, et al. w09, a novel autophagy

- enhancer, induces autophagy - dependent cell apoptosis via activation of the EGFR - mediated RAS - RAF1 - MAP2K - MAPK1/3 pathway [J]. *Autophagy*, 2017, 13(7):1093 - 1112.
- [50] SHUKLA SA, BACHIREDDY P, SCHILLING B, et al. Cancer - germline antigen expression discriminates clinical outcome to CTLA - 4 blockade [J]. *Small*, 2019, 173(4):162 - 165.
- [51] KAPRALOV A A, YANG QIN, DAR H H, et al. Redox lipid reprogramming commands susceptibility of macrophages and microglia to ferroptotic death [J]. *Nat Chem Biol*, 2020, 16(3):278 - 290.
- [52] XU C X, SUN S G, JOHNSON T, et al. The glutathione peroxidase Gpx4 prevents lipid peroxidation and ferroptosis to sustain Treg cell activation and suppression of antitumor immunity [J]. *Cell Rep*, 2021, 35(11):109235.
- [53] JIANG Q, WANG K, ZHANG X Y, et al. Platelet membrane - camouflaged magnetic nanoparticles for ferroptosis - enhanced cancer immunotherapy [J]. *Small*, 2020, 16(22):e2001704.
- [54] GUO P Y, WANG L, SHANG W T, et al. Intravesical in situ immunostimulatory gel for triple therapy of bladder cancer [J]. *ACS Appl Mater Interfaces*, 2020, 12(49):54367 - 54377.
- [55] MA S, HENSON E S, CHEN Y, et al. Ferroptosis is induced following siramesine and lapatinib treatment of breast cancer cells [J]. *Cell Death Dis*, 2016, 7(7):e2307.
- [56] CHEN J J, GALLUZZI L. Fighting resilient cancers with iron [J]. *Trends Cell Biol*, 2018, 28(2):77 - 78.
- [57] ROH J L, KIM E H, JANG H, et al. Nrf2 inhibition reverses the resistance of cisplatin - resistant head and neck cancer cells to artesunate - induced ferroptosis [J]. *Redox Biol*, 2017, 11:254 - 262.
- [58] IOANNOU N, HAGNER P R, STOKES M, et al. Triggering interferon signaling in T cells with avadomide sensitizes CLL to anti - PD - L1/PD - 1 immunotherapy [J]. *Blood*, 2021, 137(2):216 - 231.
- [59] KALBASI A, TARIVERANMOSHABAD M, HAKIMI K, et al. Uncoupling interferon signaling and antigen presentation to overcome immunotherapy resistance due to JAK1 loss in melanoma [J]. *Sci Transl Med*, 2020, 12(565):eabb0152.
- [60] GRASSO C S, TSOI J, ONYSHCHENKO M, et al. Conserved interferon - γ signaling drives clinical response to immune checkpoint blockade therapy in melanoma [J]. *Cancer Cell*, 2020, 38(4):500 - 515. e3.
- [61] ZAIDI M R, MERLINO G. The two faces of interferon - γ in cancer [J]. *Clin Cancer Res*, 2011, 17(19):6118 - 6124.
- [62] WANG W M, GREEN M, CHOI J E, et al. CD8⁺ T cells regulate tumour ferroptosis during cancer immunotherapy [J]. *Nature*, 2019, 569(7755):270 - 274.
- [63] LIAO P, WANG W M, WANG W C, et al. CD8⁺ T cells and fatty acids orchestrate tumor ferroptosis and immunity via ACSL4 [J]. *Cancer Cell*, 2022, 40(4):365 - 378. e6.
- [64] FRIEDMANN ANGELI JP, XAVIER DA SILVA TN, SCHILLING B. CD8⁺ T cells PUF(A) ing the flames of cancer ferroptotic cell death [J]. *Cancer Cell*, 2022, 40(4):346 - 348.
- [65] STOCKWELL B R, JIANG X J. A Physiological function for ferroptosis in tumor suppression by the immune system [J]. *Cell Metab*, 2019, 30(1):14 - 15.
- [66] WANG Q Y, WANG Y P, DING J J, et al. A bioorthogonal system reveals antitumour immune function of pyroptosis [J]. *Nature*, 2020, 579(7799):421 - 426.
- [67] HARATANI K, YONESAKA K, TAKAMURA S, et al. U3 - 1402 sensitizes HER3 - expressing tumors to PD - 1 blockade by immune activation [J]. *J Clin Invest*, 2020, 130(1):374 - 388.
- [68] ERKES D A, CAI W J, SANCHEZ I M, et al. Mutant BRAF and MEK inhibitors regulate the tumor immune microenvironment via pyroptosis [J]. *Cancer Discov*, 2020, 10(2):254 - 269.
- [69] ROSENBAUM S R, WILSKI N A, APLIN A E. Fueling the fire: inflammatory forms of cell death and implications for cancer immunotherapy [J]. *Cancer Discov*, 2021, 11(2):266 - 281.
- [70] CHEHRAZI - RAFFLE A, DORFF T B, PAL S K, et al. Wnt/ β - catenin signaling and immunotherapy resistance: lessons for the treatment of urothelial carcinoma [J]. *Cancers (Basel)*, 2021, 13(4):889.
- [71] CAO D, CHEN D, XIA J N, et al. Artesunate promoted anti - tumor immunity and overcame EGFR - TKI resistance in non - small - cell lung cancer by enhancing oncogenic TAZ degradation [J]. *Biomed Pharmacother*, 2022, 155:113705.
- [72] NGUYEN CDK, YI C L. YAP/TAZ signaling and resistance to cancer therapy [J]. *Trends Cancer*, 2019, 5(5):283 - 296.
- [73] MARKOWITSCH S D, SCHUPP P, LAUCKNER J, et al. Artesunate inhibits growth of sunitinib - resistant renal cell carcinoma cells through cell cycle arrest and induction of ferroptosis [J]. *Cancers (Basel)*, 2020, 12(11):3150.
- [74] ZHANG G N, ASHBY CR JR, ZHANG Y K, et al. The reversal of antineoplastic drug resistance in cancer cells by β - elemene [J]. *Chin J Cancer*, 2015, 34(11):488 - 495.
- [75] AMERIGOS DADDY J C K, CHEN M L, RAZA F, et al. Co - encapsulation of mitoxantrone and β - elemene in solid lipid nanoparticles to overcome multidrug resistance in leukemia [J]. *Pharmaceutics*, 2020, 12(2):191.
- [76] CHEN P, LI X J, ZHANG R N, et al. Combinative treatment of β - elemene and cetuximab is sensitive to KRAS mutant colorectal cancer cells by inducing ferroptosis and inhibiting epithelial - mesenchymal transformation [J]. *Theranostics*, 2020, 10(11):5107 - 5119.
- [77] GONG Z, LIU Z G, DU K Y, et al. Potential of β - elemene induced ferroptosis through Pole2 - mediated p53 and PI3K/AKT signaling in lung cancer cells [J]. *Chem Biol Interact*, 2022, 365:110088.
- [78] FENG Y X, LI C C, LIU S W, et al. β - elemene restrains pten mRNA degradation to restrain the growth of lung cancer cells via MET-TL3 - mediated N6 methyladenosine modification [J]. *J Oncol*, 2022, 2022:3472745.
- [79] 王峥嵘, 范焕芳, 张倩, 等. β - 榄香烯阻断 JAK2 - STAT3 信号通路促进紫杉醇对肺癌细胞增殖和凋亡作用研究 [J]. *中华中医药学刊*, 2019, 37(7):1600 - 1604.
- [80] WANG H Y, MA Y Y. β - Elemene alleviates cisplatin resistance in oral squamous cell carcinoma cell via inhibiting JAK2/STAT3 pathway in vitro and in vivo [J]. *Cancer Cell Int*, 2022, 22(1):244.

- [81] ZOU W W, XU S P. Galangin inhibits the cell progression and induces cell apoptosis through activating PTEN and Caspase-3 pathways in retinoblastoma[J]. *Biomed Pharmacother*, 2018, 97: 851-863.
- [82] ZOU Y, LI R, KUANG D B, et al. Galangin inhibits cholangiocarcinoma cell growth and metastasis through downregulation of MicroRNA-21 expression[J]. *Biomed Res Int*, 2020, 2020: 5846938.
- [83] 韩立卓. 高良姜素调控程序性细胞死亡配体1表达并增强T细胞活力的机制研究[D]. 吉林: 延边大学, 2022.
- [84] WU B, XU C W, DING H S, et al. Galangin inhibits neointima formation induced by vascular injury via regulating the PI3K/AKT/mTOR pathway[J]. *Food Funct*, 2022, 13(23): 12077-12092.
- [85] LI X C, JIANG J W, YANG Z F. Galangin suppresses RANKL-induced osteoclastogenesis via inhibiting MAPK and NF- κ B signaling pathways[J]. *J Cell Mol Med*, 2021, 25(11): 4988-5000.
- [86] 汪舒云, 吴霖光, 陈彬, 等. 扶正类中药调节肿瘤免疫抑制微环境的研究现状[J]. *世界科学技术-中医药现代化*, 2022, 24(10): 3862-3868.
- [87] 朱月伊, 宋运来, 石晓兰. 基于PD-1/PD-L1表达影响探讨四君子汤对NK细胞及结肠癌作用的研究[J]. *中国免疫学杂志*, 2021, 37(3): 295-300, 306.
- [88] LI J, SUN G Z, LIN H S, et al. The herb medicine formula 《Yang Wei Kang Liu》 improves the survival of late stage gastric cancer patients and induces the apoptosis of human gastric cancer cell line through Fas/Fas ligand and Bax/Bcl-2 pathways[J]. *Int Immunopharmacol*, 2008, 8(9): 1196-1206.
- [89] HUANG J M, LIU D, WANG Y W, et al. Ginseng polysaccharides alter the gut microbiota and kynurenine/tryptophan ratio, potentiating the antitumor effect of anti-programmed cell death 1/programmed cell death ligand 1 (anti-PD-1/PD-L1) immunotherapy[J]. *Gut*, 2022, 71(4): 734-745.
- [90] 李津津, 杨金颖, 孙芳芳, 等. 黄芪多糖对Lewis荷瘤小鼠PD-1/PD-L1表达的影响[J]. *天津药学*, 2022, 34(5): 9-13, 37.
- [91] 谭炜焯, 金钊, 王倩, 等. PD-1/PD-L1抑制剂的中药性能探讨[J]. *新中医*, 2020, 52(12): 206-208.
- [92] KOCH M, RENNERT J, SCHULZ C. Pancerebellitis under immunotherapy with pembrolizumab[J]. *Dtsch Arztebl Int*, 2022, 119(47): 820.
- [93] CHANG H L, KUO Y H, WU L H, et al. The extracts of *Astragalus membranaceus* overcome tumor immune tolerance by inhibition of tumor programmed cell death protein ligand-1 expression[J]. *Int J Med Sci*, 2020, 17(7): 939-945.
- [94] LI W Y, XU D N, LI B X, et al. The polysaccharide of *Atractylodes macrocephala* koidz (PAMK) alleviates cyclophosphamide-mediated immunosuppression in geese, possibly through novel_mir2 targeting of CTLA4 to upregulate the TCR-NFAT pathway[J]. *RSC Adv*, 2018, 8(47): 26837-26848.
- [95] 韩懿存, 陈玉龙, 范修琦, 等. 白术多糖通过靶向miR-34a抑制食管癌细胞免疫检查点PD-L1表达的机制研究[J]. *中国中药杂志*, 2022, 47(6): 1658-1665.
- [96] REN D M, VILLENEUVE N F, JIANG T, et al. Brusatol enhances the efficacy of chemotherapy by inhibiting the Nr1 α -mediated defense mechanism[J]. *Proc Natl Acad Sci U S A*, 2011, 108(4): 1433-1438.
- [97] WANG T, CHEN Z Y, CHEN H, et al. Brusatol inhibits the growth of renal cell carcinoma by regulating the PTEN/PI3K/AKT pathway[J]. *J Ethnopharmacol*, 2022, 288: 115020.
- [98] OH E T, KIM C W, KIM H G, et al. Brusatol-mediated inhibition of c-Myc increases HIF-1 α degradation and causes cell death in colorectal cancer under hypoxia[J]. *Theranostics*, 2017, 7(14): 3415-3431.
- [99] 陈姣, 林聃, 杨杰, 等. 中药抗肿瘤的增效减毒效应研究进展[J]. *中国科学: 生命科学*, 2022, 52(6): 920-934.
- [100] HAYAKAWA T, YAGUCHI T, KAWAKAMI Y, et al. Enhanced anti-tumor effects of the PD-1 blockade combined with a highly absorptive form of curcumin targeting STAT3[J]. *Cancer Sci*, 2020, 111(12): 4326-4335.
- [101] DAI C, ZHOU X, WANG L, et al. Rocaglamide prolonged allograft survival by inhibiting differentiation of Th1/Th17 cells in cardiac transplantation[J]. *Oxidative medicine and cellular longevity*, 2022, 2022: 2048095.
- [102] YAN X W, YAO C, FANG C, et al. Rocaglamide promotes the infiltration and antitumor immunity of NK cells by activating cGAS-STING signaling in non-small cell lung cancer[J]. *Int J Biol Sci*, 2022, 18(2): 585-598.
- [103] YAO C, NI Z Y, GONG C Y, et al. Rocaglamide enhances NK cell-mediated killing of non-small cell lung cancer cells by inhibiting autophagy[J]. *Autophagy*, 2018, 14(10): 1831-1844.
- [104] 黄婉怡. 米仔兰活性成分辣酰胺与抗PD-1抗体协同抗肿瘤的作用及其分子免疫机制[D]. 上海: 上海中医药大学, 2020.
- [105] CHEN C, ZHOU J, JI C Y. Quercetin: a potential drug to reverse multidrug resistance[J]. *Life sciences*, 2010, 87(11-12): 333-338.
- [106] JING L, LIN J R, YANG Y, et al. Quercetin inhibiting the PD-1/PD-L1 interaction for immune-enhancing cancer chemopreventive agent[J]. *Phytother Res*, 2021, 35(11): 6441-6451.
- [107] LV J, JIA Y T, LI J, et al. Gegen Qinlian decoction enhances the effect of PD-1 blockade in colorectal cancer with microsatellite stability by remodelling the gut microbiota and the tumour microenvironment[J]. *Cell Death Dis*, 2019, 10(6): 415.
- [108] 吕骥. 葛根苓连汤联合PD-1抑制剂通过重塑免疫微环境治疗结肠癌的机制研究[D]. 石家庄: 河北医科大学, 2020.
- [109] 王楠, 方兴刚, 廖莎, 等. 葛根苓连汤基于Wnt/ β -catenin信号通路对结肠癌模型大鼠肠道菌群作用机制研究[J]. *辽宁中医药大学学报*, 2023, 25(2): 49-53.
- [110] 李晓靖, 朱文伟, 缪正炆, 等. 健脾滋肾泻火方通过LMP2/JAK/STAT信号通路改善原发免疫性血小板减少症的机制研究[J]. *中药药理与临床*, 2023, 39(7): 10-14.
- [111] 杨小静. 健脾滋肾法对SLE患者血清补体及补体调节蛋白CD55和CD59水平影响的临床研究[D]. 合肥: 安徽中医药大学, 2016.